Please fill in only headlines in each objective in the table, and the full description in the explanatory notes section

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| Objective | Beginning  | Ending  | Note  |
| Examine the pore resealing time in reversible electroporated gram-**negative bacteria** in a medium containing a **fluorescent dye.** | 1.1.22 | 31.2.22 | Two monthsA suspension of***Pseudomonas putida*** in PBS (with different conductivities: 1 and 155–4058 µS cm-1) will be exposed to PEF treatment in a moderate electric field (1-4 kV cm-1) and variable physical parameters (e.g., pulse number, current density, and total specific energy). Immediately following PEF treatment, the suspension will be diluted in brain heart infusion (BHI), a rich medium and including the fluorescent dye Lucifer Yellow (LY). **LY-positive cells** will be immediately analyzed using a **flow cytometer**, which will continue until the number of LY-positive cells will be reduced to near zero. **Controls: (1)** Identical conditions, but the bacteria will be suspended in ultra-pure (UP) water**; (2)** non PEF-treated ***P. putida*** will be diluted as in the experiment and will be examined for LY-positive cells. |
| Modeling: Basic kinetic model for mass transfer into the cell. | 1.1.22 | 31.6.22 | Six monthsDevelop a basic kinetic model for mass transfer into the bacterial cell and preliminary experimental data validation.Estimate the relevant transport properties.  |
| Modeling: Basic CFD model for PEF treatment.  | 1.1.22 | 31.6.22 | Six monthsDevelop a basic CFD model for field parameters during PEF exposure.Numerical description of the PEF process. |
| Examine pore size and resealing time in reversible electroporated gram-**negative bacteria** in a medium containing different **relative hydrophilic compounds.** | 1.3.22 | 31.10.22 | Eight monthsA suspension of***P. putida*** in PBS (with different conductivities: 1 and 155–4058 µS cm-1) will be exposed to PEF treatment in a moderate electric field (1-4 kV cm-1) and variable physical parameters (e.g., pulse number, current density, and total specific energy). Immediately following PEF treatment, the suspension will be diluted into the nutrient-rich BHI medium containing different relative **hydrophilic compounds** including **phenol, bisphenol A, ellagic acid, epigallocatechin gallate, procyanidin B2, and theaflavin-3-gallate,** with molecular weights of 94.11- 716.604 g/mol. Permeabilization rates of the hydrophilic compounds will be analyzed using High Performance Liquid Chromatography (HPLC) at different time intervals (for example, every 15 min) immediately after BHI dilution until the permeabilization rate is reduced to zero.**Controls: (1)** Identical conditions, but bacteria will be suspended in UP water**; (2)** non PEF-treated ***P. putida*** will be diluted as in the experiment, and the permeabilization rateof **hydrophilic compounds** will be examined.  |
| Examine the pore size and resealing time in reversible electroporated gram-**negative bacteria** in a medium containing **different hydrophobic compounds.** | 1.11.21 | 31.5.22 | Eight monthsA suspension of ***P. putida*** in PBS (with different conductivities: 1 and 155–4058 µS cm-1) will be exposed to PEF treatment in a moderate electric field (1-4 kV cm-1) and variable physical parameters (e.g., pulse number, current density, and total specific energy). Immediately after PEF treatment, the suspension will be diluted into the nutrient-rich BHI medium containing **hydrophobic compounds** from one aromatic hydrocarbon to 10 rings, which include seven aromatic hydrocarbon rings and three rings of five carbons. Compounds tested include **benzene, naphthalene, anthracene, pyrene, benzo[e]pyrene, and decacyclene**, with molecular weights of 78.12- 450.5 g/mol. It is important to note that each examined molecule's concentration will be lower than the concentration that causes cell damage. The hydrophobic compounds' permeabilization rate will be analyzed using HPLC at different time intervals (for example, every 15 min) immediately after the dilution in BHI until the permeabilization rate is reduced to zero.**Controls: 1-** Identical conditions, but the bacteria will be suspended in UP water**; 2-** non PEF-treated ***P. putida*** will be diluted as in the experiment, and the rate permeabilization of the **hydrophobic compounds** will be examined.  |
| Examine the pore resealing time in reversible electroporated gram-**positive bacteria** in a medium containing a **fluorescent dye.** | 1.6.22 | 31.7.22 | Two monthsA suspension *of* ***Staphylococcus* *aureus*** in PBS (with different conductivities: 1 and 155–4058 µS cm-1) will be exposed to PEF treatment in a moderate electric field (1-4 kV cm-1) and variable physical parameters (e.g., pulse number, current density, and total specific energy). Immediately after PEF treatment, the suspension will be diluted into the nutrient-rich BHI medium containing the fluorescent dye, LY. The **LY-positive cells** will be analyzed using a **flow cytometer** immediately after dilution and continue until the number of LY-positive cells is reduced to near zero. **Controls: (1)** Identicalconditions, but the bacteria will be suspended in UP water**; (2)** non PEF-treated *S. aureus* will be diluted and examined for LY-positive cells identically as in the experimental group.  |
| Examine pore size and resealing time in reversible electroporated gram-**positive bacteria** in a medium containing relative **different hydrophilic compounds.** | 1.8.22 | 31.3.23 | Eight monthsA suspension of***S. aureus*** in PBS (with different conductivities: 1 and 155–4058 µS cm-1) will be exposed to PEF treatment in a moderate electric field (1-4 kV cm-1) and variable physical parameters (e.g., pulse number, current density, and total specific energy). Immediately after PEF treatment, the suspension will be diluted into the nutrient-rich BHI medium containing relative different **hydrophilic compounds,** such as **phenol, bisphenol A, ellagic acid, epigallocatechin gallate, procyanidin B2, and theaflavin-3-gallate** with molecular weights of 94.11- 716.604 g/mol. The permeabilization rate of the hydrophilic compounds will be analyzed using High Performance Liquid Chromatography (HPLC) at different time intervals (for example, every 15 min) immediately after dilution in BHI until the permeabilization rate is reduced to zero.**Controls: (1)** Identical conditions, but the bacteria will be suspended in UP water**; (2)** non PEF-treated ***S. aureus*** will be diluted as in the experimental group, and the rate permeabilization of **hydrophilic compounds** will be examined.  |
| Examine pore size and resealing time in reversible electroporated gram-**positive bacteria** in a medium containing **different hydrophobic compounds.** | 1.4.23 | 31.11.23 | Eight monthsA suspension of***S. aureus*** in PBS (with different conductivities: 1 and 155–4058 µS cm-1) will be exposed to PEF treatment in a moderate electric field (1-4 kV cm-1) and variable physical parameters (e.g., pulse number, current density, and total specific energy). Immediately after PEF treatment, the suspension will be diluted into the nutrient- rich BHI medium containing **hydrophobic compounds** from one aromatic hydrocarbon to 10 rings, including seven aromatic hydrocarbon rings and three rings of five carbons. Compounds include **benzene, naphthalene, anthracene, pyrene, benzo[e]pyrene, and decacyclene**, with molecular weights of 78.12- 450.5 g/mol. It is important to note that each molecule's concentration will be lower than the concentration that causes cell damage. The permeabilization rate of the hydrophobic compounds will be analyzed using HPLC at different time intervals (for example, every 15 min) immediately after the dilution in BHI until the permeabilization rate is reduced to zero.**Controls: (1)** Identical conditions, but the bacteria will be suspended in UP water**; (2)** non PEF-treated ***S. aureus*** will be diluted as in the experimental group, and the rate permeabilization of **hydrophobic compounds** will be examined.  |
| Modeling: Membrane dynamics study: Kinetic and CFD Modeling.  | 1.7.22 | 31.12.23 | Twelve monthsDevelopment of a model for cell membrane destruction and recovery with experimental data validation. For the tested cell and tracers, identification of the relevant mechanisms and drive forces on the membrane.  |
| Bacterial protoplast preparation. | 1.12.23 | 31.2.24 | Three months*P. putida* and *S. aureus* peptidoglycans will be digested employing murein hydrolases, and the commonly hen egg-white lysozyme. Since the outer membrane of gram-negative bacteria prevents enzyme entry, these types of cells require pre-treatment with a chelating agent (e.g., EDTA) or detergent (e.g., Triton X-100) for outer membrane removal. |
| Examine the pore resealing time in reversible electroporated gram-**bacterial protoplast** in a medium containing a **fluorescent dye** | 1.3.24 | 31.5.24 | Three monthsA suspension of ***bacterial protoplast*** in PBS with different **selected** conductivities appropriate for protoplasts: 1 and 155–4058 µS cm-1) will be exposed to PEF treatment in a moderate electric field (1-4 kV cm-1) and variable **selected** physical parameters (e.g., pulse number, current density, and total specific energy). After PEF treatment, the suspension will be diluted into the nutirent rich BHI medium containing the fluorescent dye, LY. The **LY-positive protoplast bacteria** will be immediately analyzed using a **flow cytometer** and will continue until the LY-positive protoplast cells are reduced to near zero. **Controls: N**on PEF-treated bacterial protoplastwill be diluted as in the experimental group and examined for LY-positive cells.  |
| Examine pore size and resealing time in reversible electroporated **bacterial protoplast** in a medium containing a selected relative **hydrophilic compound** | 1.6.24 | 31.10.24 | Five monthsA suspension of**bacterial protoplast** in PBS (with different **selected** conductivities appropriate for protoplast: 1 and 155–4058 µS cm-1) will be exposed to PEF treatment in a moderate electric field (1-4 kV cm-1) and variable **selected** physical parameters (e.g., pulse number, current density, and total specific energy). Immediately after PEF treatment, the suspension will be diluted into the nutrient-rich BHI medium containing **selected hydrophilic compounds (**one of the following**:** **benzene, naphthalene, anthracene, pyrene, benzo[e]pyrene, and decacyclene**, with molecular weights of 78.12- 450.5 g/mol). The permeabilization rate of the **selected** hydrophilic compound will be analyzed using HPLC at different time intervals (for example, every 15 min) immediately dilution until the permeabilization rate is reduced to zero.**Controls:** Non PEF-treated bacterial protoplastwill be diluted as in the experimental group, and the permeabilization rate of the **hydrophilic compound** will be examined.  |
| Examine pore size and resealing time in reversible electroporated **bacterial protoplast** in a medium containing a selected **hydrophobic compound.** | 1.11.24 | 31.3.25 | Five monthsA suspension of**bacterial protoplast** in PBS (with different **selected** conductivities which is appropriate for protoplast: 1 and 155–4058 µS cm-1) will be exposed to PEF treatment in a moderate electric field (1-4 kV cm-1) and variable **selected** physical parameters (e.g., pulse number, current density, and total specific energy). Immediately after PEF treatment, the suspension will be diluted into the nutrient-rich BHI medium containing a **hydrophobic compound (**one of the following**:** **phenol, bisphenol A, ellagic acid, epigallocatechin gallate, procyanidin B2, and theaflavin-3-gallate** with molecular weights of 94.11- 716.604 g/mol). The permeabilization rate of the **selected** hydrophilic compound will be analyzed using HPLC at different time intervals (for example, every 15 min) immediately after the dilution in BHI until the permeabilization rate is reduced to zero.**Controls:** Non PEF-treated bacterial protoplastwill be diluted as in the experimental group, and the permeabilization rate of the **hydrophilic compound** will be examined.  |
| Shed light on the electroporated recovery process of *P. putida* and *S. aureus*the bacterial proteome of the PEF-treated and untreated will be examind using mass spectrometry analysis (MS).  | 1.4.25 | 31.12.25 | NIne monthsAt selected times, electroporated bacteria and control, non-treated bacteria will be collected and centrifuged. The proteins from the washed sediment will be sonicated and treated with urea, ammonium bicarbonate, and DTT. Mass spectrometry analysis will be performed at the Smoler Proteomics Center at the Technion, Israel. MS will be performed by an Q-Exactive HFX mass spectrometer (Thermo) in a positive mode using repetitively full MS scans, followed by collision-induced dissociation (HCD) of the 30 most dominant ions selected from the first MS scan. The data will be quantified by label-free analysis using the MaxQuant software 1.5.2.8, based on extracted ion currents (XICs) of peptides, enabling quantitation from each LC/MS run for each peptide identified in any of the experiments. |
| Modeling: Full kinetic model for pore size dynamics.  | 1.1.24 | 31.12.25 | Twelve monthsDevelopment of a full kinetic model for membrane destruction and recovery and experimental data validation.Simulation of cell membrane dynamics.  |
| Modeling: Full CFD model for PEF treatment and recovery stages. | 1.1.24 | 31.12.25 | Twelve monthsDevelopment of a full CFD model for field parameters during PEF treatment and recovery and experimental data validation.Numerical simulation of the PEF process on single cells and cell populations (field description). |